

Isoprenoids

What are isoprenoids? (also called terpenoids)

Definition: *hydrocarbons* structurally based on multiple isoprene units

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Introduction:

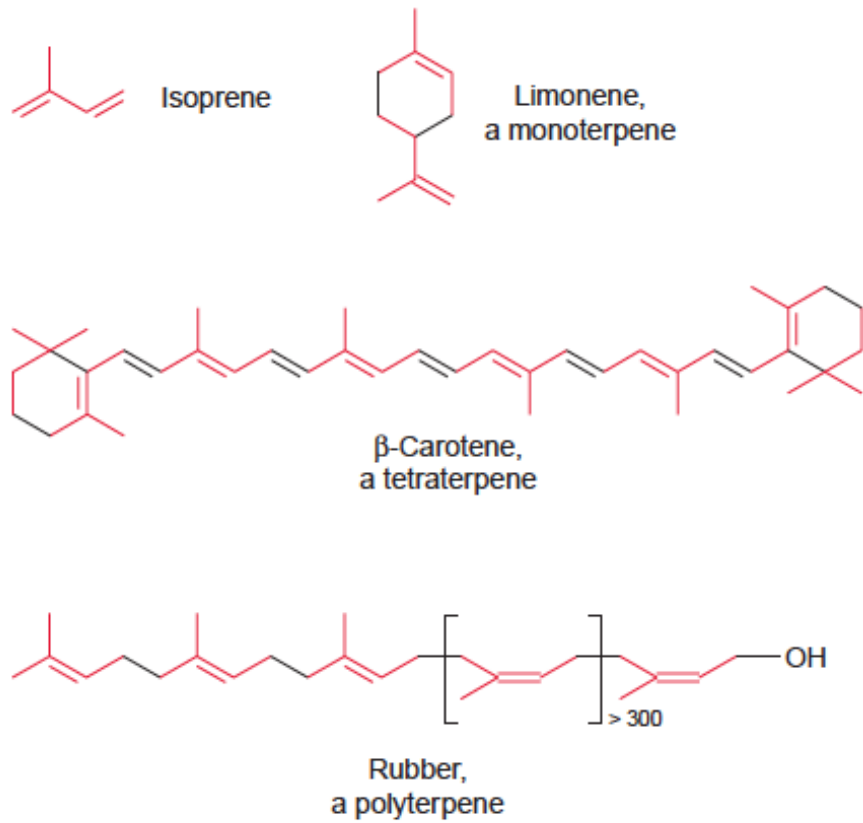
- names: 5C isoprene derivation = terpenoids (like "turpentine")
- huge structural diversity (20,000 structures), largest group of secondary plant metabolites
- exist mostly as multiples of **5 carbons units** (the Lego principle)
- types: **hemi, mono-, sesqui-, di-, tri-, tetra-terpenes** (5, 10, 15, 20, 30, 40 carbons)
 - also polyterpenes (very large)
- two complementary biosynthetic routes are known, for different groups of isoprenoids
 - i) **mevalonate** (MVA) pathway
 - ii) **deoxyxylulose phosphate** pathway

- impressive **functional diversity**: many **volatile** compounds, signals, toxins, hormones and more.
- some compounds form this family function as primary metabolites, but most fall into the secondary metabolite category
- many human uses of isoprenoids from or in plants (fragrances, flavors, pharmaceuticals, rubber)

Strategy and Outline

- learn basic biosynthesis pathways of isoprenoid building blocks.
- examples with specific functions of each class of isoprenoids

Introduction: Representative isoprenoid structures



Introduction: Functional diversity of isoprenoids

Table 17.1: Isoprenoids of higher plants

Precursor	Class	Example	Function
C ₅ : Dimethylallyl-PP	Hemiterpene	Isoprene	Protection of the photosynthetic apparatus against heat
Isopentenyl-PP		Side chain of cytokinin	Growth regulator
C ₁₀ : Geranyl-PP	Monoterpene	Pinene	Defense substance attractant
		Linalool	
C ₁₅ : Farnesyl-PP	Sesquiterpene	Capsidiol	Phytoalexin
C ₂₀ : Geranylgeranyl-PP	Diterpene	Gibberellin	Plant hormone
		Phorbol	Defense substance
		Casbene	Phytoalexin
C ₃₀ : 2 Farnesyl-PP	Triterpene	Cholesterol	Membrane constituents
		Sitosterol	
C ₄₀ : 2 Geranylgeranyl-PP	Tetraterpene	Carotenoids	Photosynthesis pigments
<i>n</i> Geranylgeranyl-PP or <i>n</i> Farnesyl-PP	Polyprenols	Prenylated proteins	Regulation of cell growth
		Prenylation of plastoquinone, ubiquinone, chlorophyll, cyt <i>a</i>	Membrane solubility of photosynthesis pigments and electron transport carriers
		Dolichols	Glucosyl carrier
		Rubber	

Overview of Isoprenoid Biosynthesis

- two complementary biosynthetic routes are known, for different groups of isoprenoids
 - i) **mevalonate** (MVA) pathway
 - ii) **deoxyxylulose phosphate** pathway

i. Mevalonate pathway

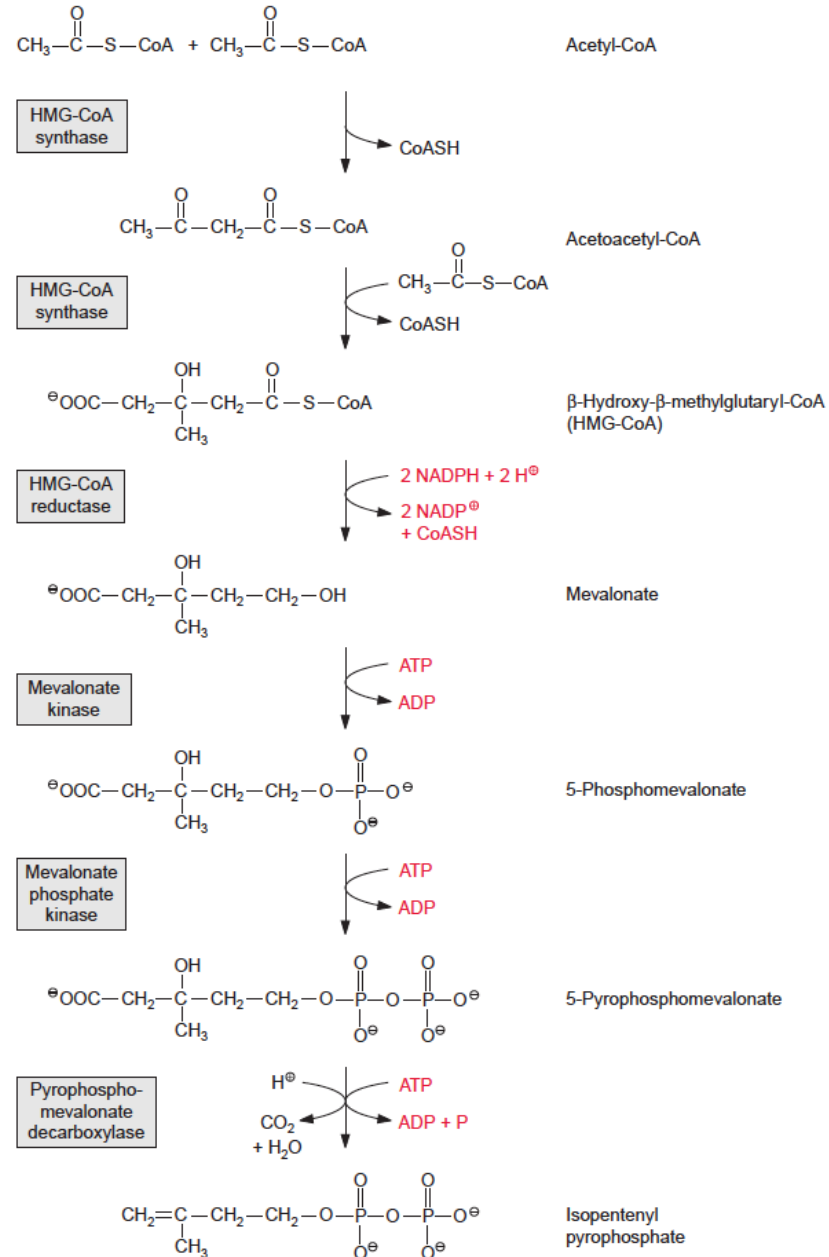
→ produces **isopentenyl pyrophosphate (IPP)**
for sesqui- and tri-terpenes

Hydroxymethylglutaryl CoA (HMGCoA)
synthase (key gateway enzyme)

- hydroxymethylglutaryl CoA (HMGCoA)
reductase

- kinases, dehydratases

- IPP isomerase makes methylallyl
pyrophosphate (DMAPP)



Deoxyxylulose phosphate (DXP) [Methylerythriol (MEP) pathway]

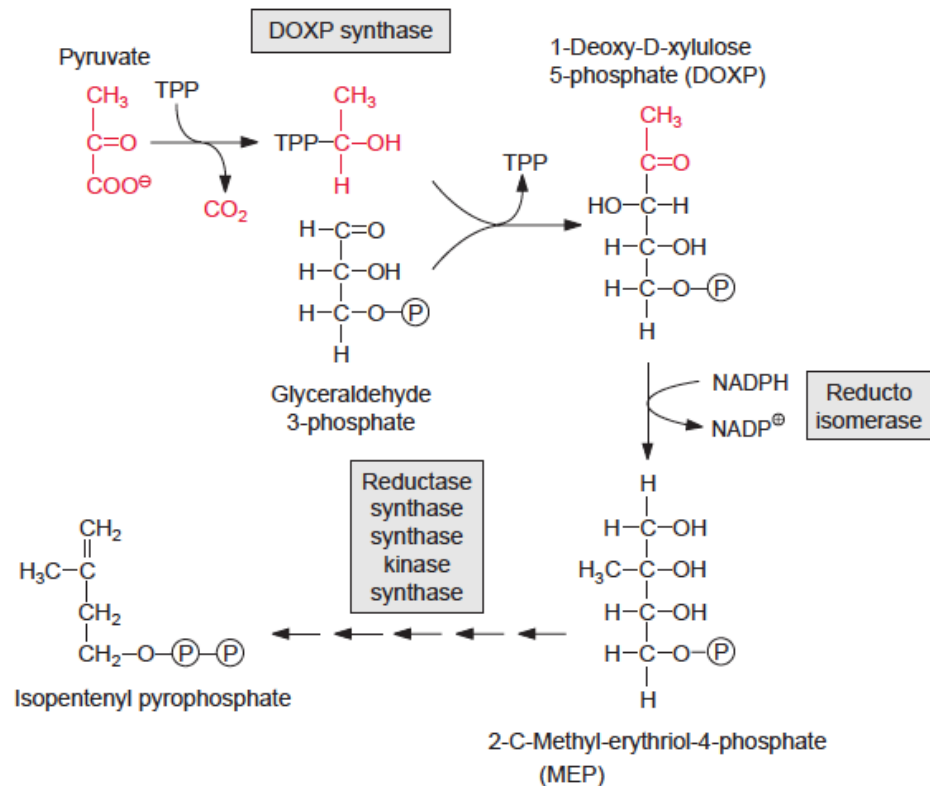
→ produces isopentenyl pyrophosphate (IPP)

ii). Deoxyxylulose phosphate (DXP) pathway

- in **plastids**,
- leads to mono-, di-, and tetra-terpenes
- also found in bacteria, some protists, algae

Precursors: **pyruvate & glyceraldehyde-3-P**

- products: both IPP and DMAPP
- key enzyme: *deoxyxylulose phosphate (DXP) synthase*



Step 2: Successive condensations of **IPP** with **DMAPP** to form larger isoprenoids

- head-to-tail ('lego')

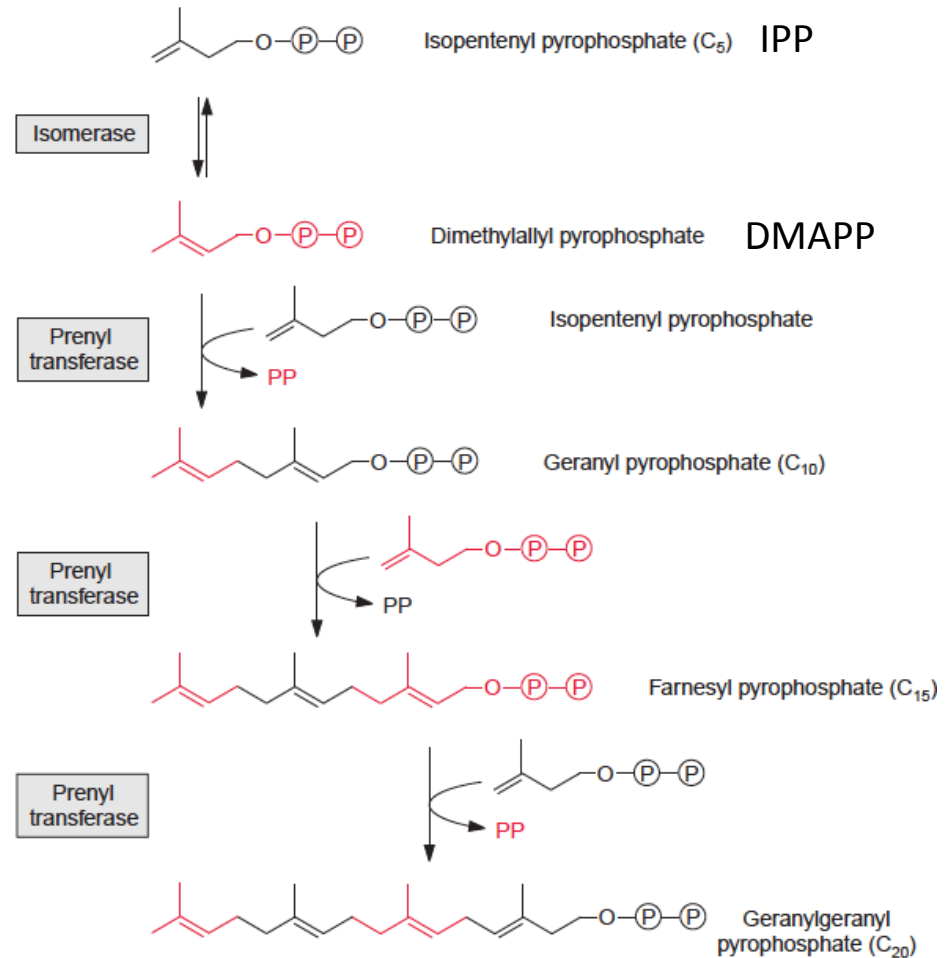
- enzymes: *prenyl transferases*

Note: one enzyme does multiple condensations, but is specific for each end product

1 IPP + 1 DMAPP --> **geranyl-PP**
geranyl-PP synthase

2 IPP + 1 DMAPP --> **farnesyl-PP**
farnesyl-PP synthase (one enzyme)

3 IPP + 1 DMAPP --> **geranylgeranyl-PP**
geranylgeranyl-PP synthase (one enzyme)

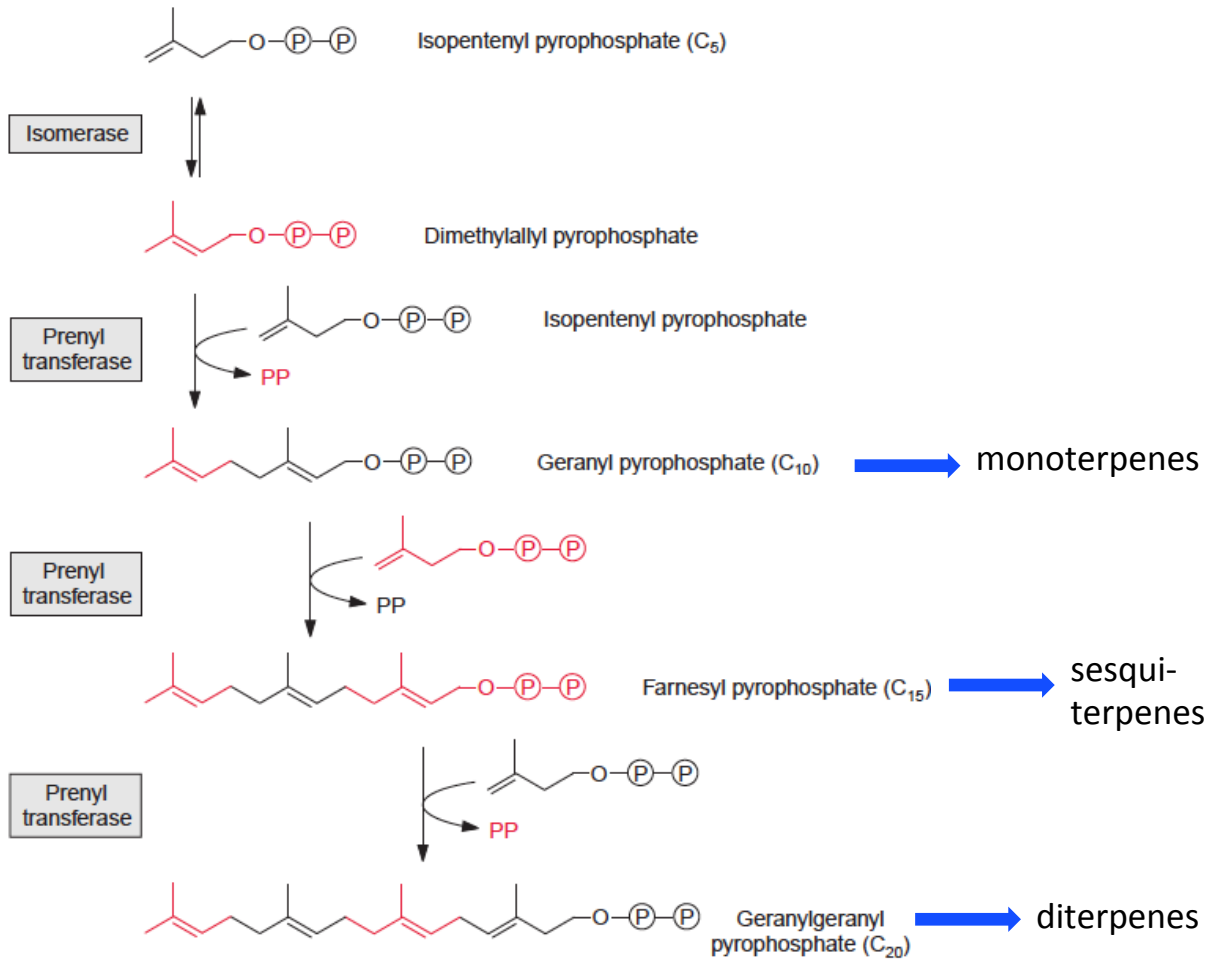


Step 2: Successive condensations of IPP with DMAPP to form larger isoprenoids

- head-to-tail ('lego')

- enzymes: *prenyl transferases*

Note: one enzyme does multiple condensations, but is specific for each end product



Step 3: Further elaborations and modifications - monoterpenes

- cyclizations (*terpene synthases - TPS*)
- hydroxylations, oxidations, reductions, rearrangements

Example of monoterpene synthases:

- **linalool synthase** (specific)
- **cineol synthase** ('sloppy')
- > multiproduct enzyme

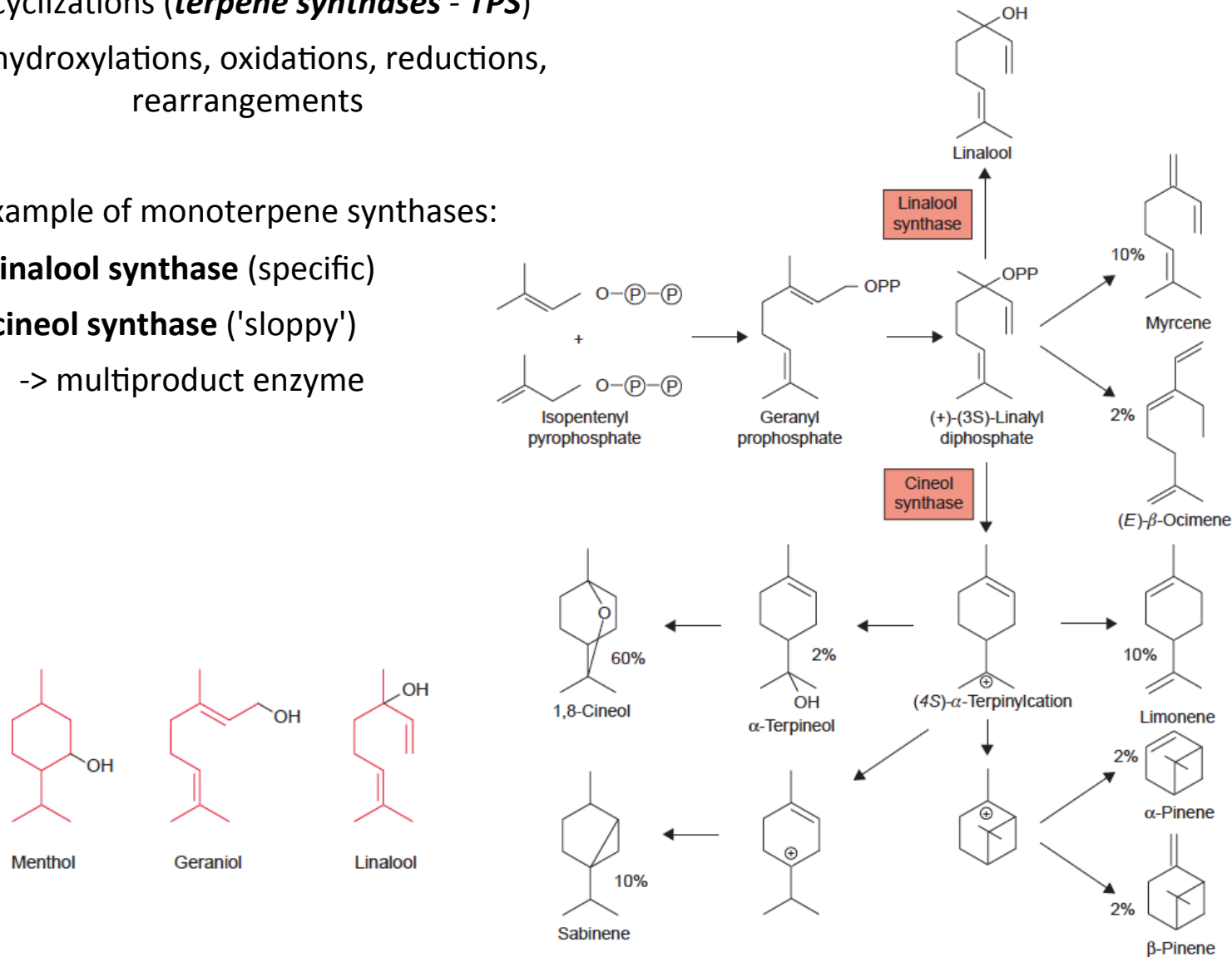


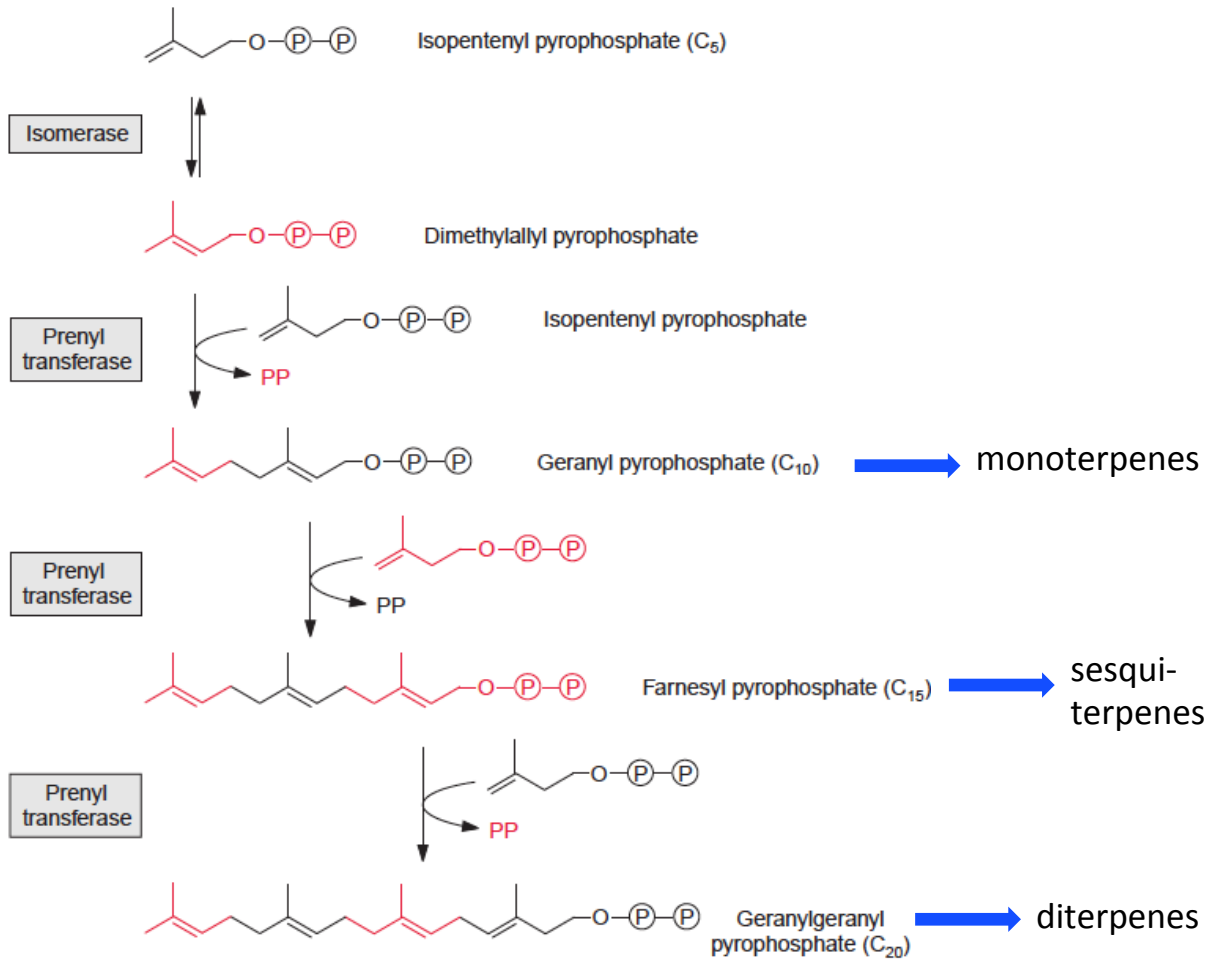
Fig 17.8, 9

Step 2: Successive condensations of IPP with DMAPP to form larger isoprenoids

- head-to-tail ('lego')

- enzymes: *prenyl transferases*

Note: one enzyme does multiple condensations, but is specific for each end product



Step 3: Further elaborations and modifications – sesquiterpene synthase (cyclase)

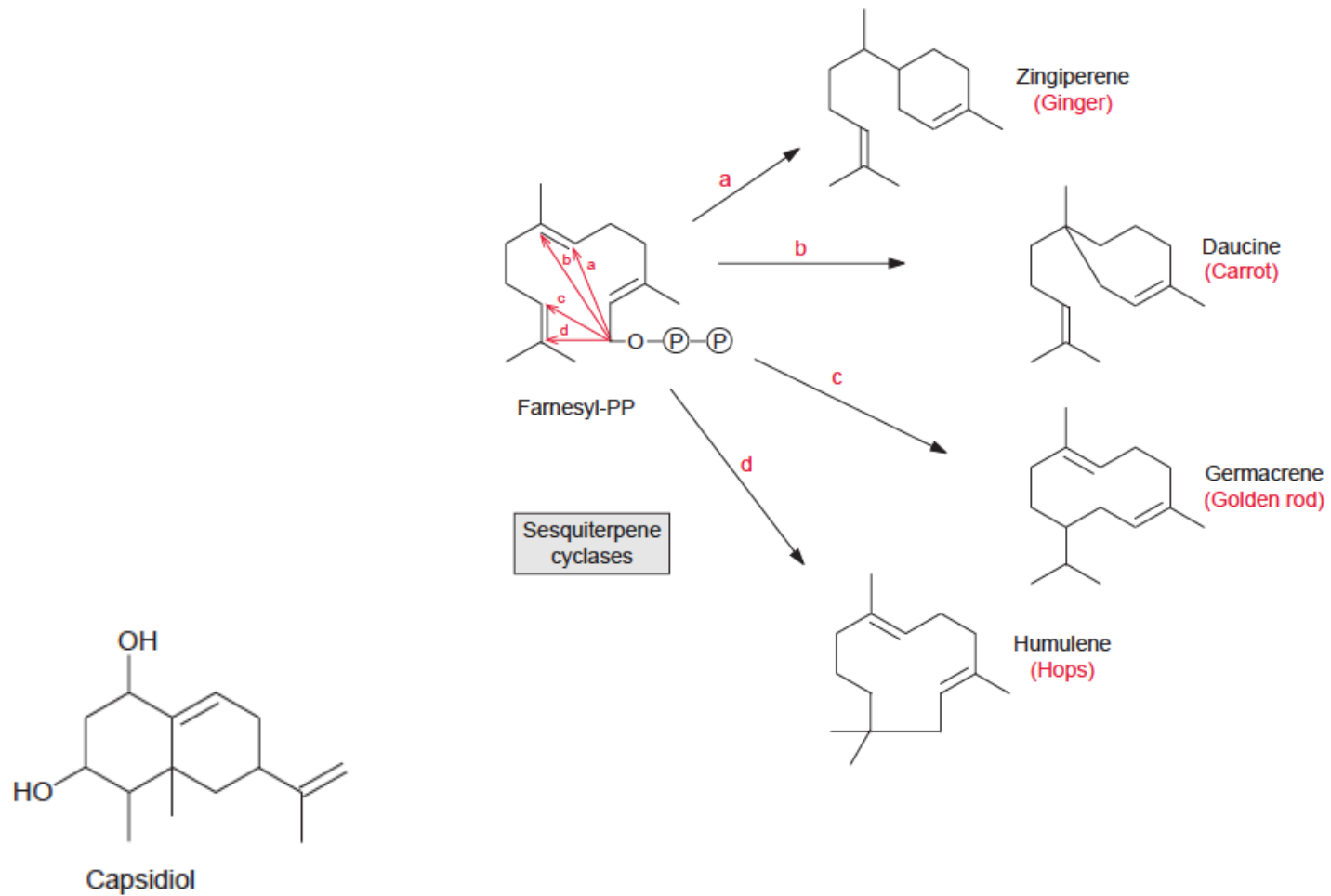
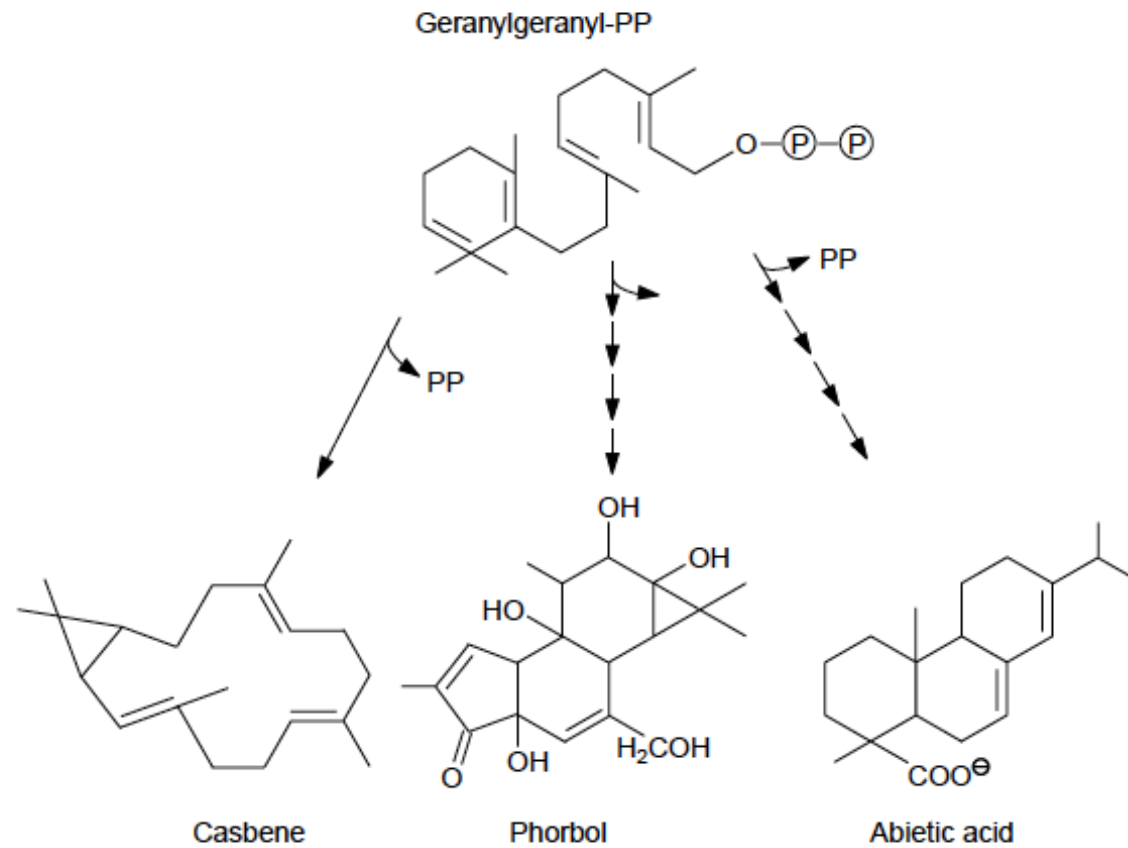


Fig 17.10

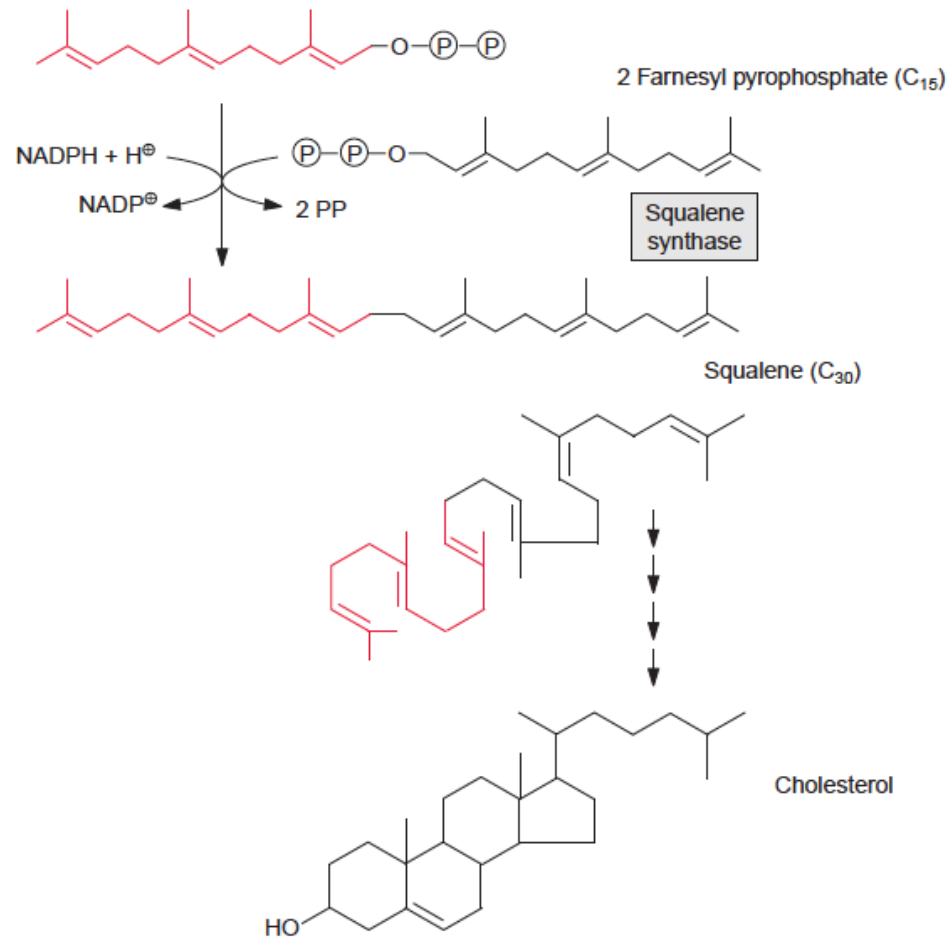
Step 3: Further elaborations and modifications - diterpenes

Fig 17.13



Step 4: Further condensations to triterpenes

Fig 17.11

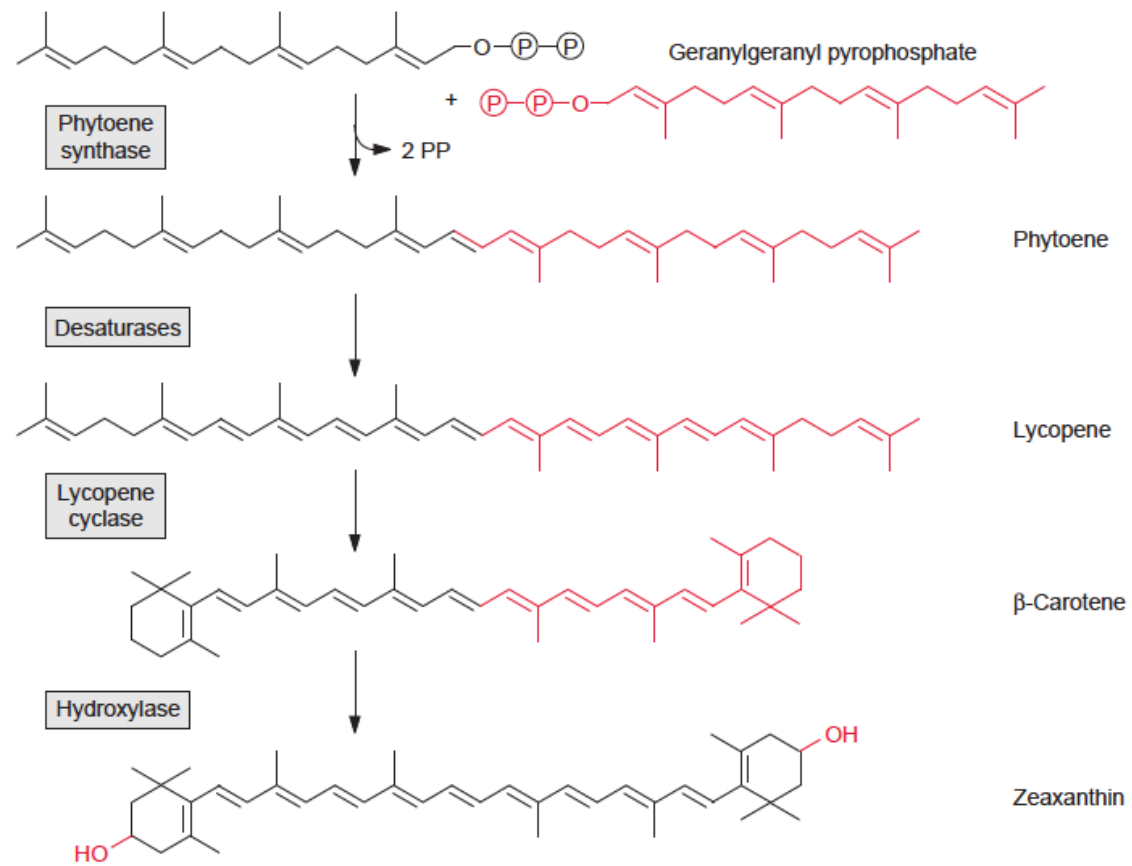


2 x farnesyl-PP → triterpenes (sterols and steroids, cardiac glycosides, others)

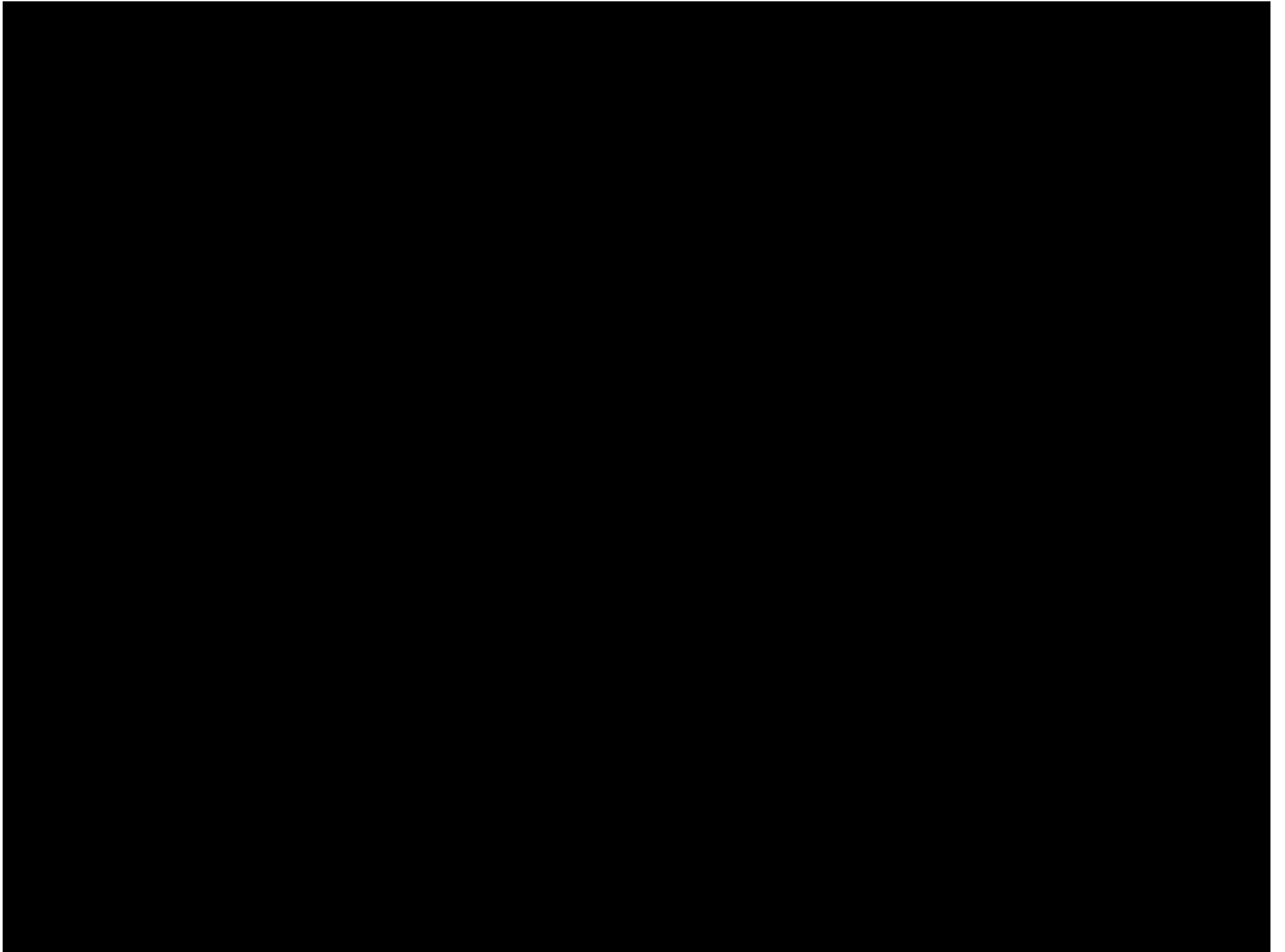
key enzyme: squalene synthase

Step 3: Further condensations - tetraterpenes

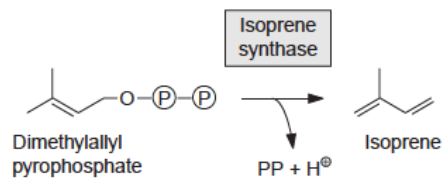
Fig 17.14



2 x geranylgeranyl-PP → tetraterpenes (carotenoids, xanthophylls and derived products)
enzyme: phytoene synthase

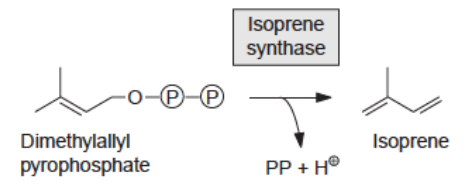
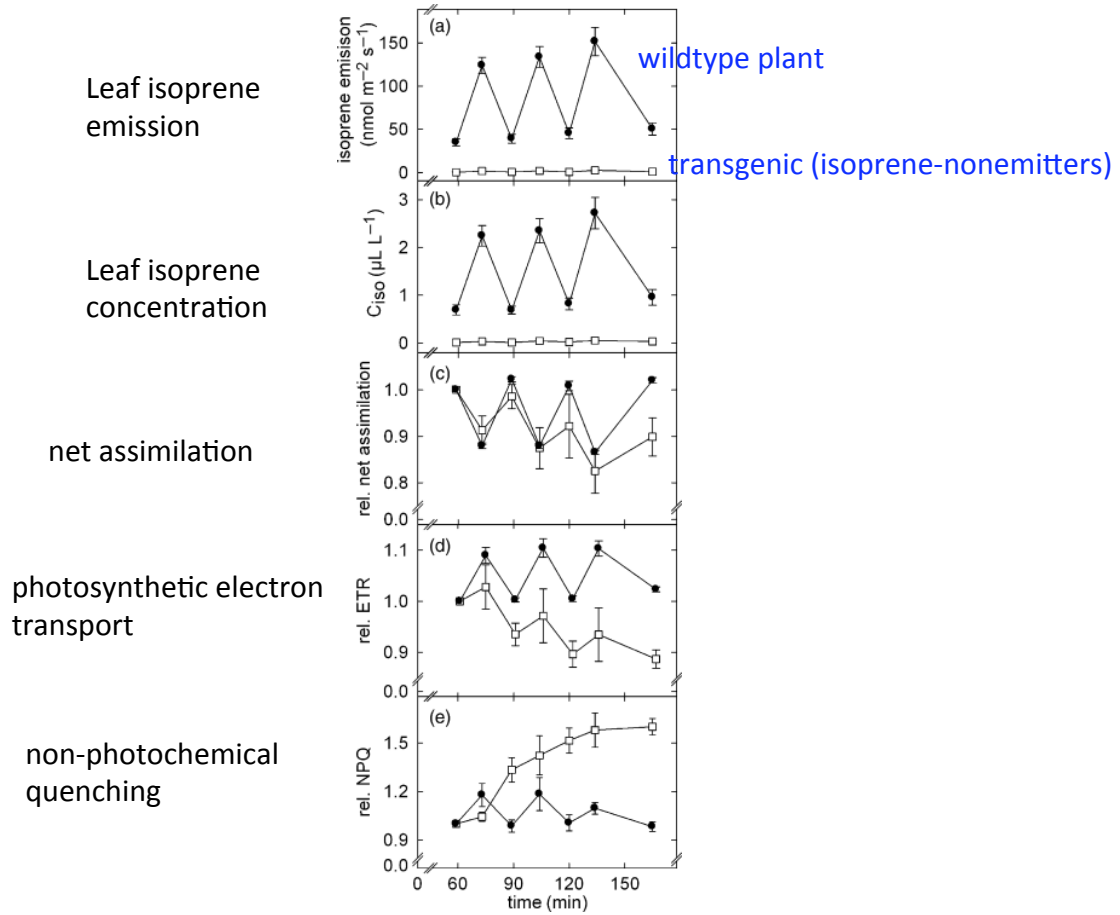


Hemiterpenes: Isoprene emission from leaves of trees (heat-stressed)



www.atmos.colostate.edu/~heald/research.html

Transgenic (non-isoprene emitting) poplar plants are more heat-sensitive



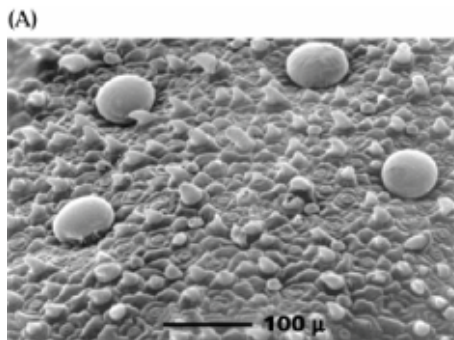
2. Functions for monoterpenes and sesquiterpenes

i) direct defenses (toxic/repellent)

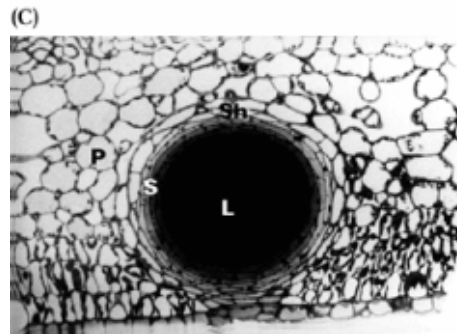
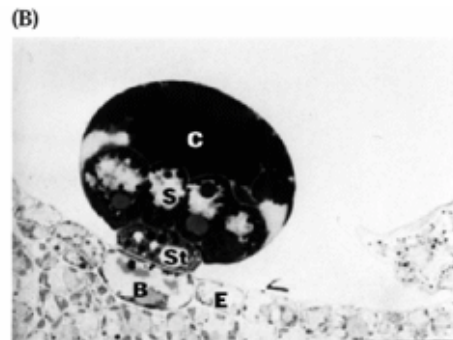
Peppermint trichomes (special modified hairs) contain monoterpenes and sesquiterpenes
 - released by herbivores (insects) breaking trichome

Structures that accumulate (lipophilic) monoterpenes

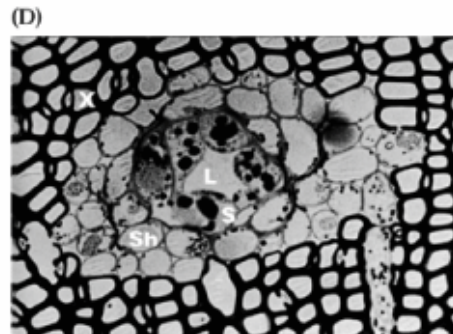
glands (thyme)



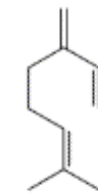
glandular trichomes (spearmint)



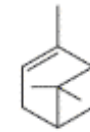
secretory cavity (lemon leaf)



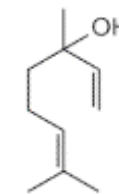
resin duct (Jeffrey pine)



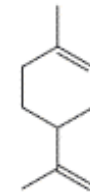
Myrcene



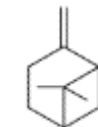
α -Pinene



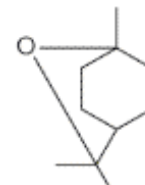
Linalool



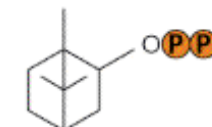
Limonene



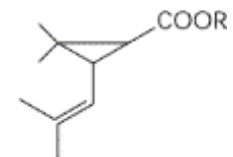
β -Pinene



1,8-Cineole



Bornyl diphosphate



Pyrethrin I

iii) Many mono- or sesquiterpenes are **volatile** (= *ecological signals*)

- attract pollinators, seed dispersors (see scents and fragrances
(eg) linalool, limonene (recall: methyl benzoate, a phenolic volatile)
- attract **predators** (=act as **indirect defenses**)



www2.unine.ch/entomo/page24332_en_US.html

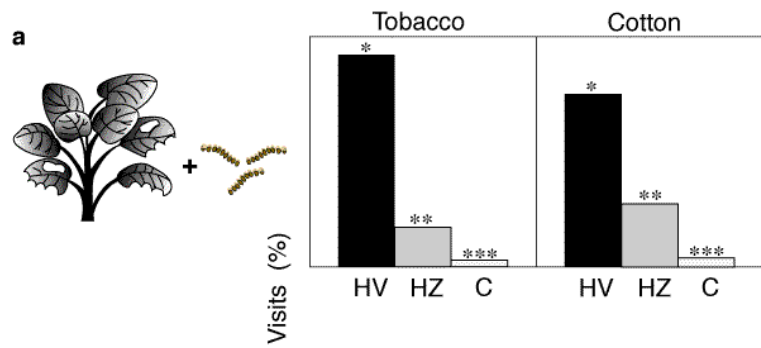
Parasitic wasps attacking Spodoptera on maize leaf



Parasitoid pupae
on lepidopteran pest

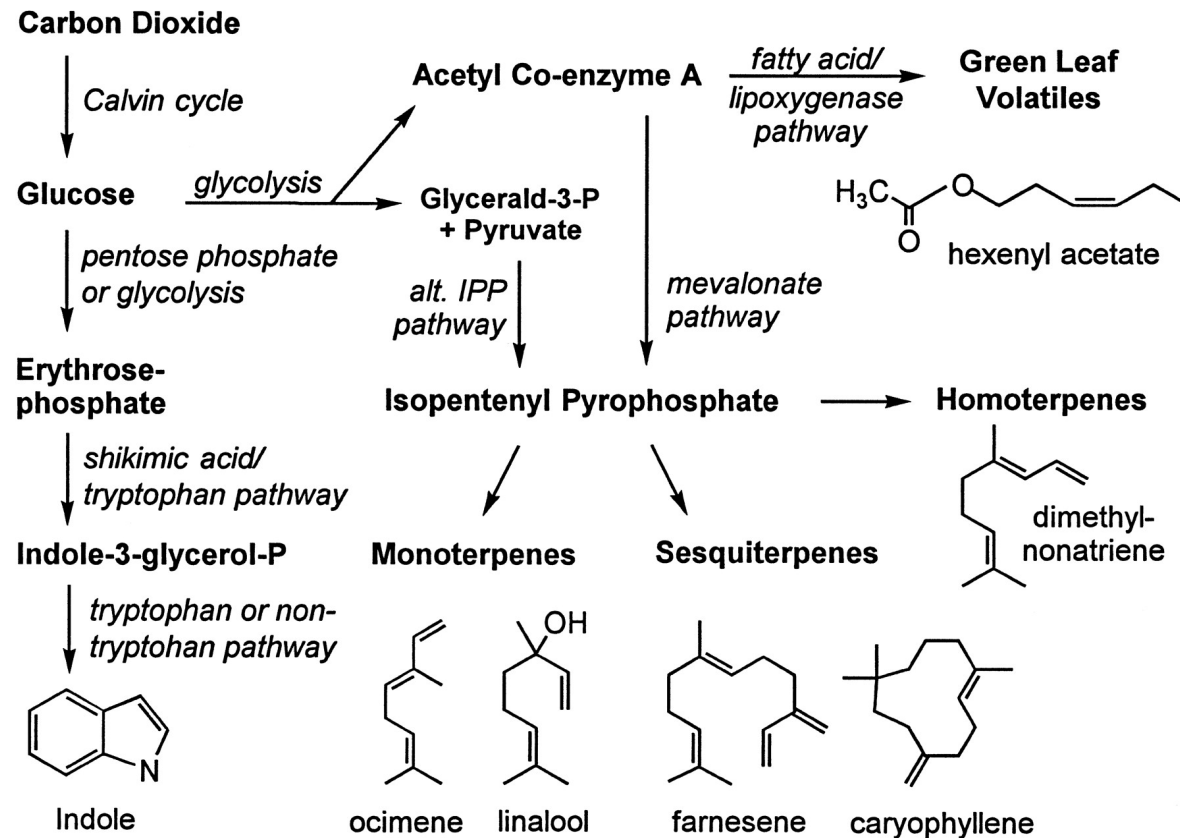
Mono- and sesquiterpenes: volatile cues for parasitic wasps and predators

Cardiochiles nigriceps (parasitic wasp) searching for *Heliothis virescens* (caterpillar)



(De Moraes et al, Nature 393: 570)

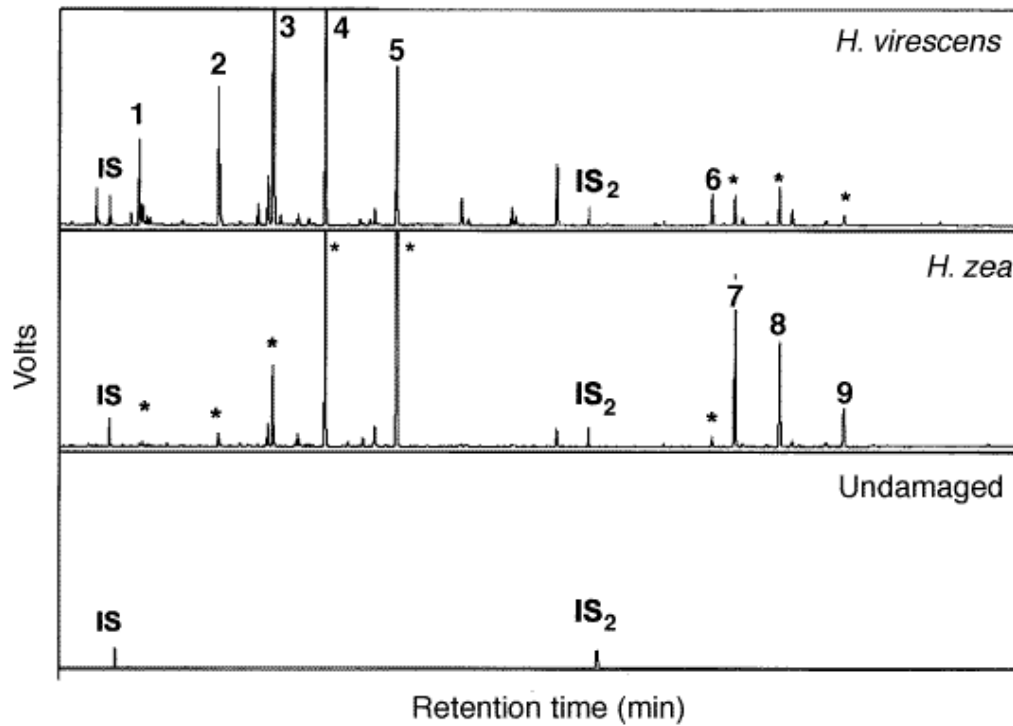
Biosynthetic pathways leading to the release of plant volatiles



Par P. W., Tumlinson J. H. Plant Physiol. 2010:121:325-332

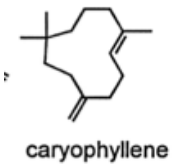
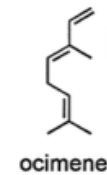


Gas chromatograph comparing damage by two types of caterpillars

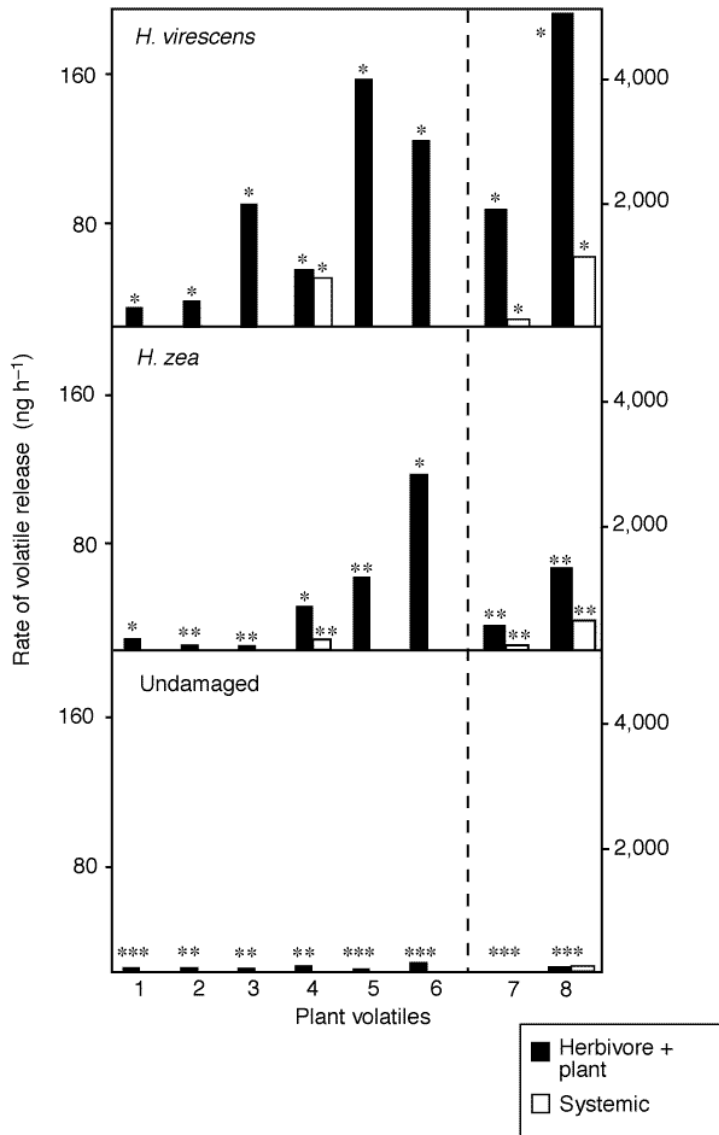


(De Moraes et al, Nature 393: 570)

- (7) β -Ocimene
- (8) α -Ocimene
- (9) β -caryophyllene



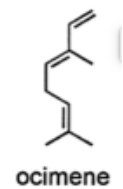
Gas chromatograph comparing damage by two types of caterpillars



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Nature 393: 570)

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(8) α-Ocimene



*What is the basis of differences in release of volatiles from *H. zea* and *H. virescens*?*

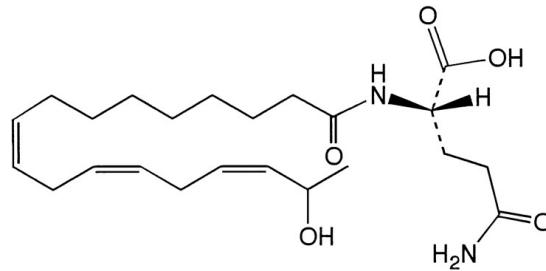
-> the biochemistry of caterpillar regurgitant or "spit"....



An Elicitor of Plant Volatiles from Beet Armyworm Oral Secretion

H. T. Alborn, T. C. J. Turlings,* T. H. Jones, G. Stenhagen
J. H. Loughrin,† J. H. Tumlinson‡

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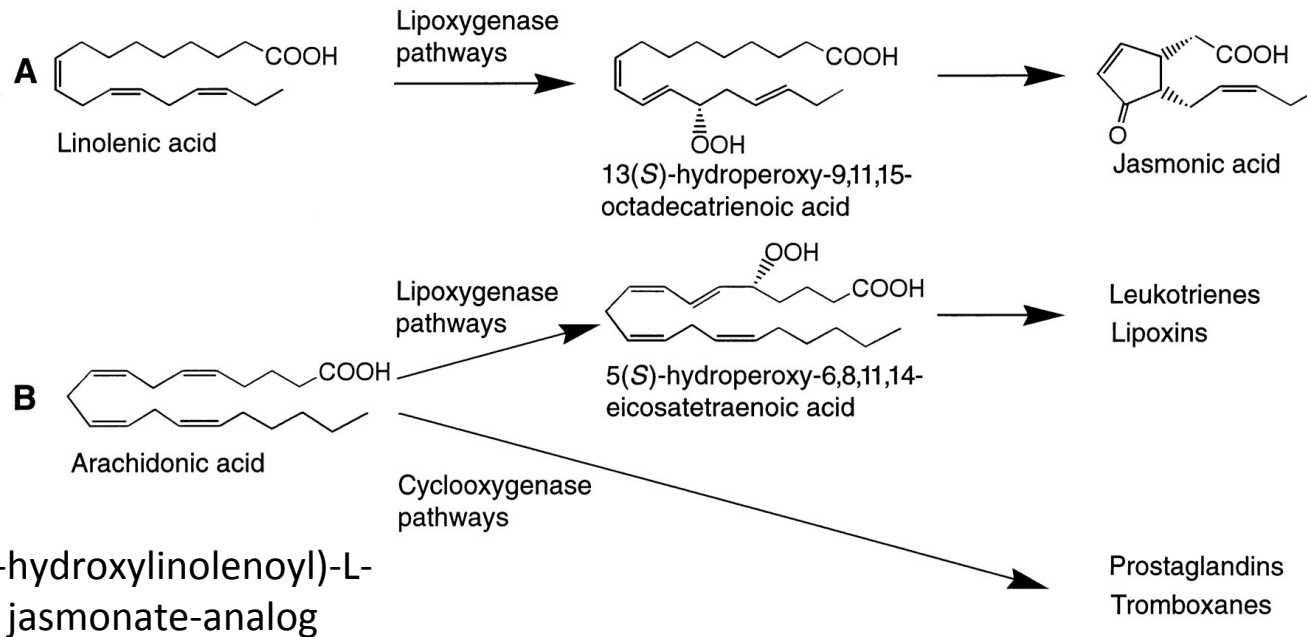
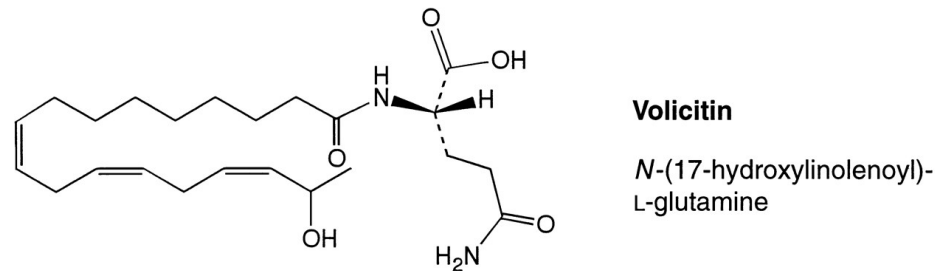
Volicitin

N-(17-hydroxylinolenoyl)-
L-glutamine

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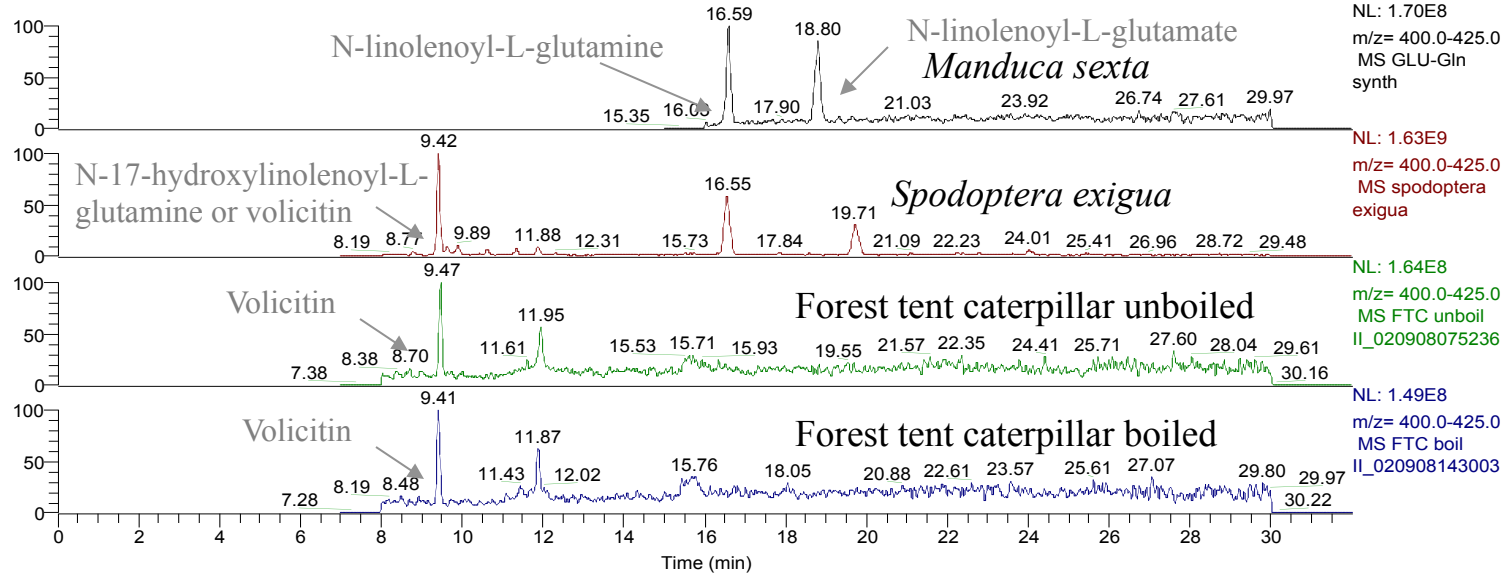


Alborn et al, Science 276: 945 (1997)

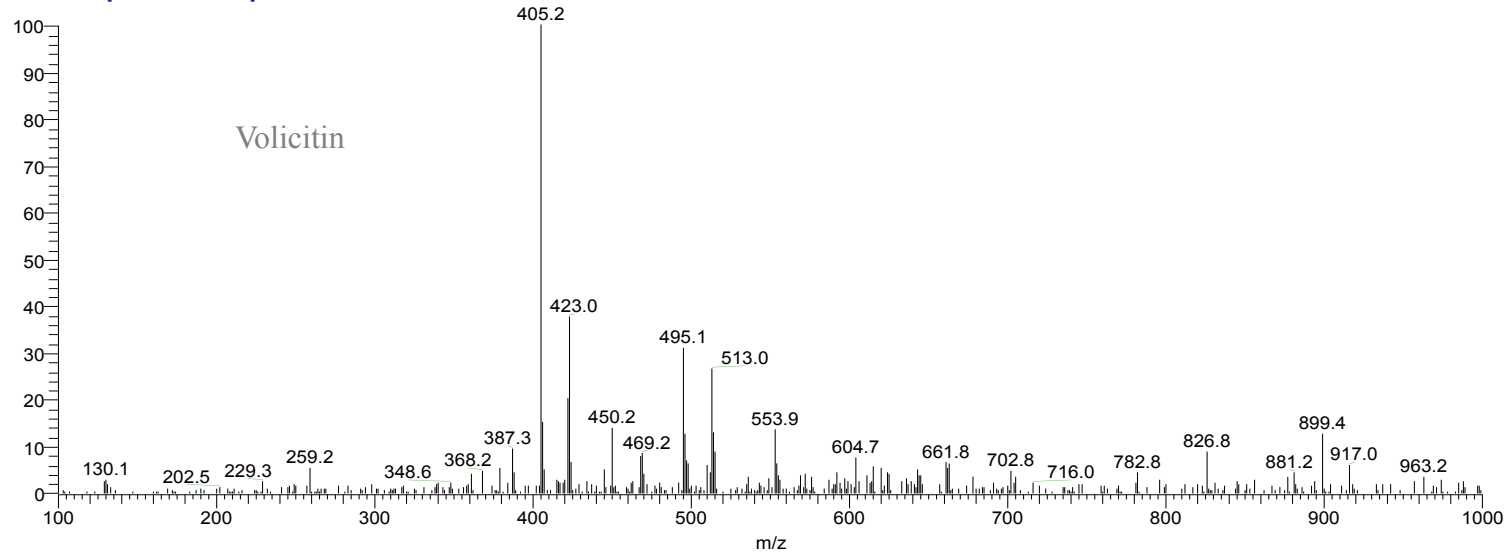
The biochemistry of caterpillar spit



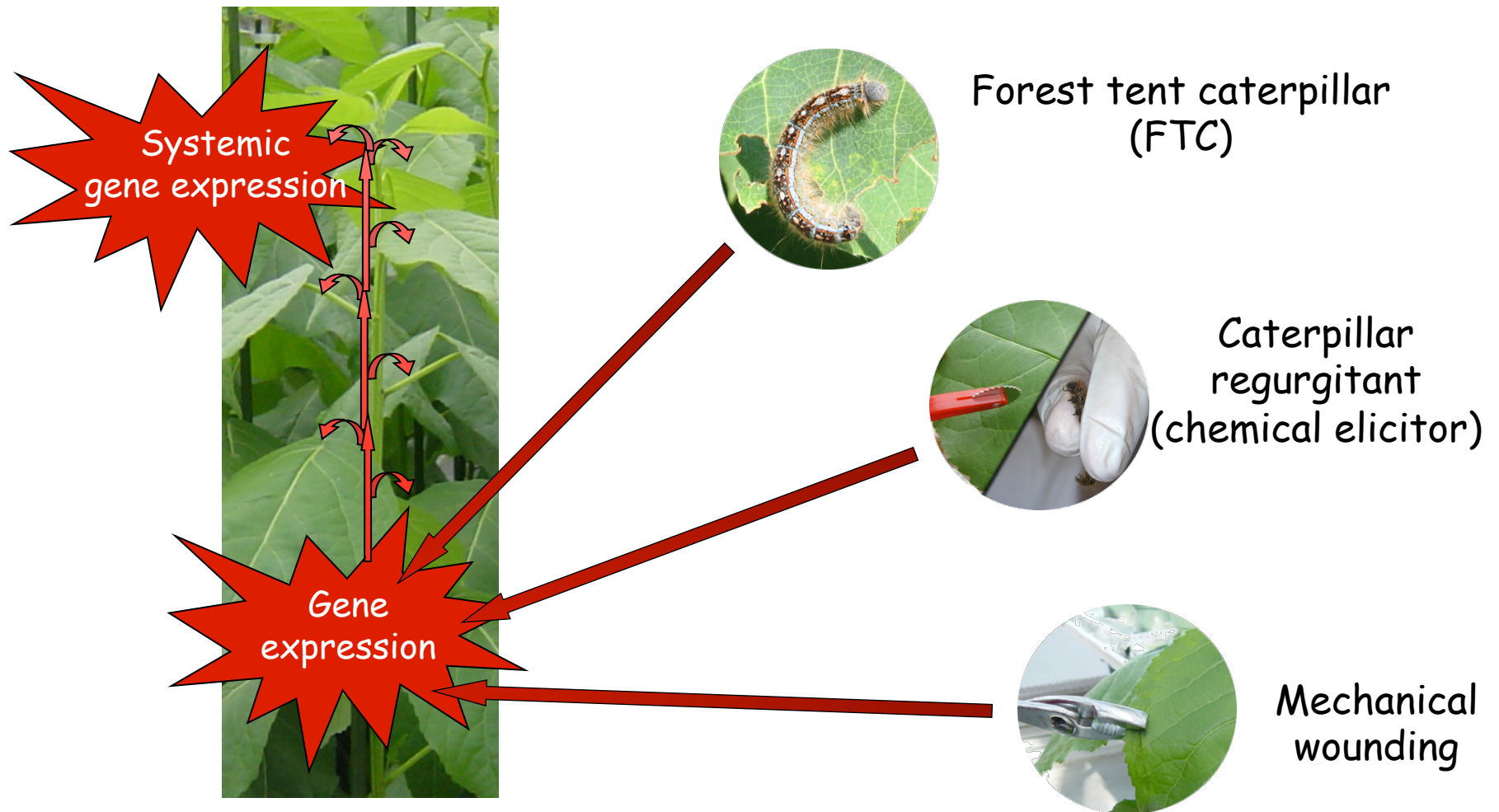
RT: 0.00 - 32.00 SM: 7G



spodoptera exigua #163 RT: 9.42 AV: 1 SB: 2 9.15, 9.66 NL: 8.92E8
T: + c Full ms [100.00-1000.00]

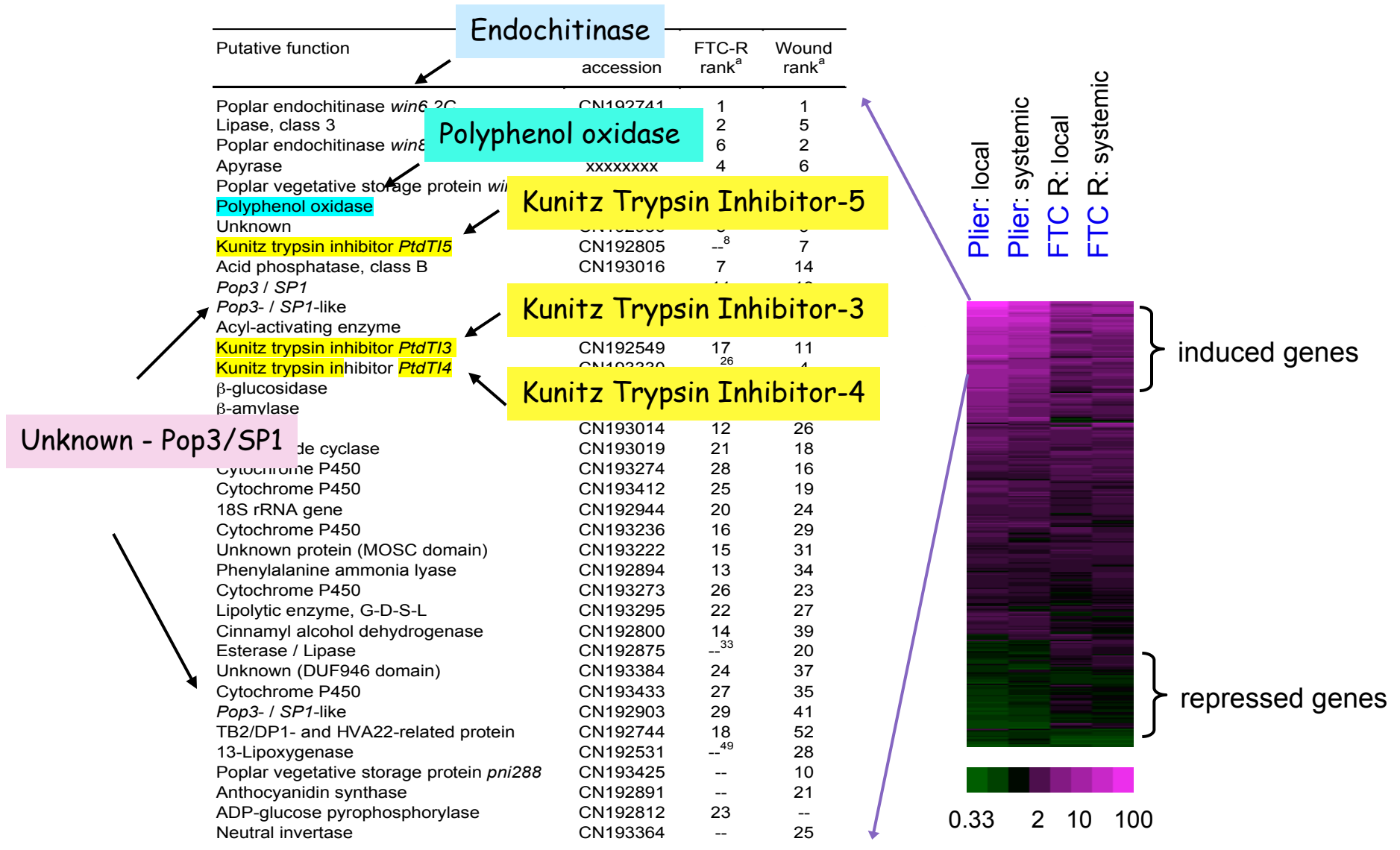


Defense of poplar against insect herbivory is an *active process* and can be triggered by *multiple signals*



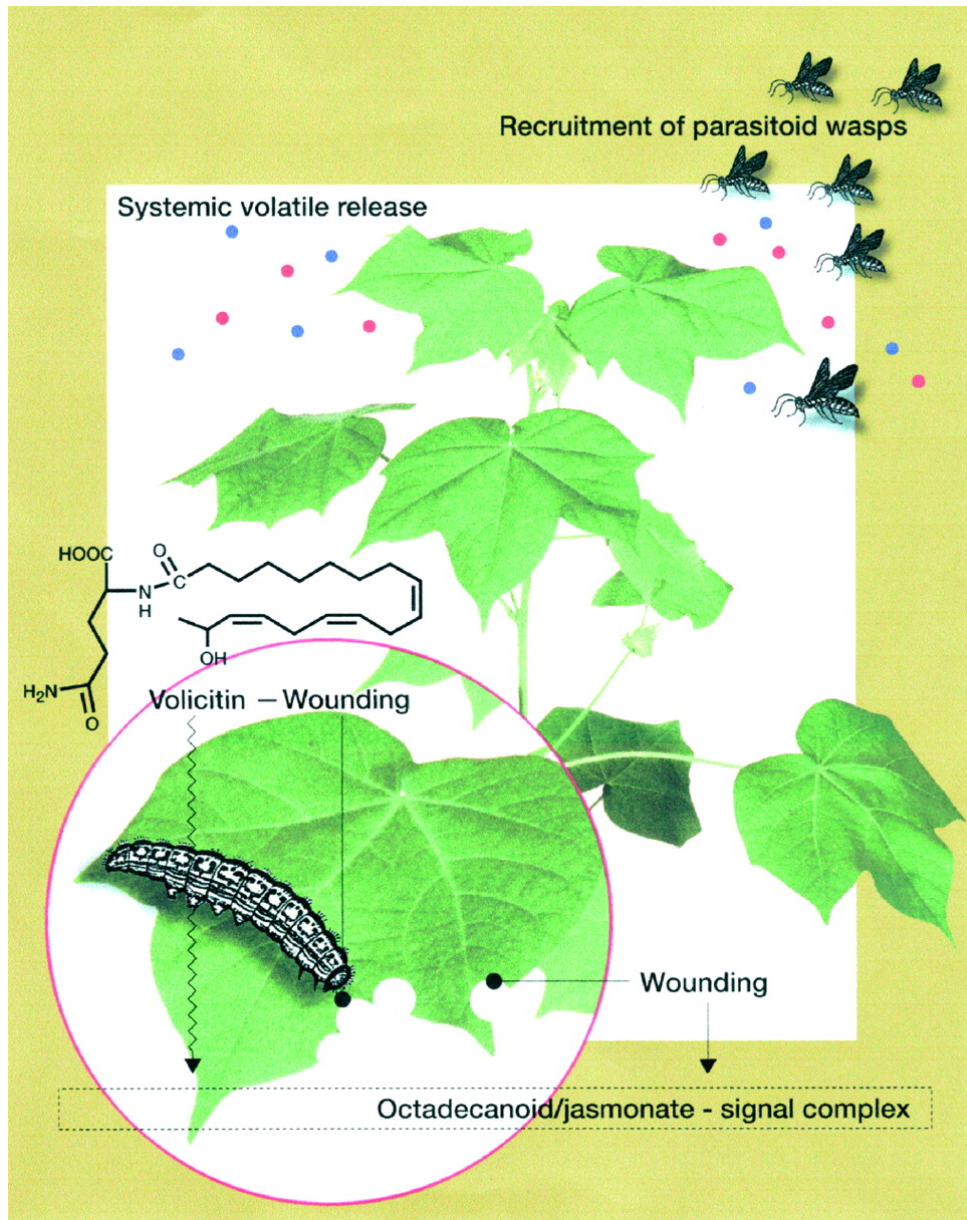
Macroarray analysis of the poplar herbivore defense response

What kinds of genes are induced? (2-fold, $P < 0.05$, 117 total)



Major and Constabel, New Phytol 172: 617-635 (2006)

Increase of volatile compounds released by plants in response to insect feeding



Par P. W., Tumlinson J. H. Plant Physiol. 1999:121:325-332

