## HOF GROUP PROGRESS REPORTS

Undergrads complete at end of co-op, summer, 498, or 499 work term.
Grad students and post-docs write an original report due JULY 1 AND JAN 1

Suggested outline (modify as needed)

## Introduction

Background
Objectives
WHO CARES
Results (last six months only)
Experimental plan
What approaches have you tried (full synthetic schemes)
What has worked
WHAT HASN'T WORKED

## Discussion

Significance of results
Future experiments/New objectives
Experimental section
General methods (solvent purification, commercial sources, instrumentation, etc.)
Procedure and characterization for each compound - see next page

## References

printed in JACS format using Endnote - if you don't know how, ask someone.
Appendix
Printouts of all new ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra, including integrals and peak labels

A few extra requirements:
-Use Symbol font for all greek characters ( $\delta, \Delta, \alpha$, etc.)
-Use double spacing throughout.
-Start all Chemdraw files by selecting File... Apply Document Settings From... ACS
Document 1996. Make sure you use fixed bond lengths and angles.
-Synthetic schemes should include conditions and yields by the arrow for each reaction, or in the caption directly below the scheme.
-ALL compounds are numbered sequentially in the order in which they appear in the text.
Use these numbers in the text, in the figures, and in the experimental section.

## EXPERIMENTAL DATA IS REQUIRED FOR EVERY NEW COMPOUND

## Checklist:

Mp (if solid)
$\square$ IR (text only)
$\square{ }^{1} \mathrm{H}$ NMR (text form and printout)
$\square{ }^{13} \mathrm{C}$ NMR (text form and printout)
$\square$ High-Res MS (text form)

## Notes:

Write a complete experimental for every NEW compound you make to a reasonable degree of purity ( $\sim 95 \%$ ). Follow the example below, making sure you have all $J$ values, multiplicities, integrations, punctuation, and italics in the format indicated. Give ${ }^{1} \mathrm{H}$ assignments where possible.

Compounds that have been fully characterized in the literature need only have a ${ }^{1} \mathrm{H}$ NMR in text and printout form and a reference to the original preparation/characterization.

NMR spectra must be transferred to the group computer and given proper peak labels and integrals AS THEY ARE COLLECTED. Your files on the group computer must be organized in such a way that they make sense to future group members.

Put your printouts of NMR, MS, and IR data in a binder organized by notebook number.
Printouts of ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMRs including peak labels and integrals must be labeled for easy identification and included as an appendix for each progress report.

## Sample Experimental:

Compound (( $\pm$ )-29c).


A mixture of aryl bromide ( $\pm$ )-28 ( $66 \mathrm{mg}, 0.109 \mathrm{mmol}), \mathrm{Pd}(\mathrm{OAc})_{2}(1.2 \mathrm{mg}, 0.005 \mathrm{mmol}), 2-$
(dicyclohexylphosphino)biphenyl ( $4.0 \mathrm{mg}, 0.011 \mathrm{mmol}$ ), 2-methoxybenzeneboronic acid (30.0
$\mathrm{mg}, 0.19 \mathrm{mmol})$, and $\mathrm{K}_{3} \mathrm{PO}_{4}(69.4 \mathrm{mg}, 0.327 \mathrm{mmol})$ was placed in an oven-dried flask and flushed with argon for 20 min . Anhydrous DMF ( 2.0 ml ) was added, and the reaction was heated at $100^{\circ} \mathrm{C}$ overnight. After cooling to $22^{\circ} \mathrm{C}$, the reaction mixture was filtered through silica and concentrated to dryness on a rotary evaporator. Column chromatography (4:1 hexanes/AcOEt) gave the product ( $51.2 \mathrm{mg}, 75 \%$ ) as a white solid. $\mathrm{Mp} 45-48^{\circ} \mathrm{C}$. IR: $2929 w, 2857 w, 1697 \mathrm{~m}$, $2250 w, 1697 m, 1597 w, 1499 w, 1464 m, 1422 m, 1366 m, 1302 m, 1240 m, 1162 m, 1146 s, 1086 m$, $1057 w, 1027 w .{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right): \delta 0.85-0.92(m, 3 \mathrm{H}, \mathrm{H}-\mathrm{C}(14)) ; 1.16-1.37$ ( $m, 6$ H, H-C(11-13)); 1.41 ( $s, 9 \mathrm{H}, \mathrm{Boc})$; 1.49-1.75 ( $m, 4 \mathrm{H}, \mathrm{H}-\mathrm{C}(10)$, H-C(5) endo, H-C(6) exo); 1.83-1.95 ( $\mathrm{m}, 1 \mathrm{H}, \mathrm{H}-\mathrm{C}(5)$ exo); 2.44 (ddd, $J=3.2,8.6,11.8,1 \mathrm{H}, \mathrm{H}-\mathrm{C}(6)$ endo); $2.68(t, J=7.7$, $2 \mathrm{H}, \mathrm{H}-\mathrm{C}(9))$; $3.26(d t, J=1.6,4.5,1 \mathrm{H}, \mathrm{H}-\mathrm{C}(2)) ; 3.35(d, J=2.6,1 \mathrm{H}, \mathrm{H}-\mathrm{C}(3)) ; 3.61(d, J=$ 13.1, $1 \mathrm{H}, \mathrm{NCH}_{2}$ naph. $) ; 3.76\left(d, J=13.1,1 \mathrm{H}, \mathrm{NCH}_{2}\right.$ naph. $) ; 3.81\left(s, 3 \mathrm{H}, \mathrm{OCH}_{3}\right) ; 4.29(d, J=$ 5.1, $1 \mathrm{H}, \mathrm{H}-\mathrm{C}(4)) ; 4.44(t, J=4.3,1 \mathrm{H}, \mathrm{H}-\mathrm{C}(1)) ; 6.97-7.09(m, 3 \mathrm{H}, \mathrm{ArH}) ; 7.23(d, J=8.3,2 \mathrm{H}$, H-C(8)); 7.25-7.36 (m, $4 \mathrm{H}, \mathrm{ArH}) ; 7.42$ ( $t d, J=1.4,7.7,1 \mathrm{H}, \mathrm{ArH}) ; 7.72$ ( $d, J=8.3,2 \mathrm{H}, \mathrm{H}-$ $\mathrm{C}(7)) .{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right): \delta 14.2 ; 22.7 ; 24.6 ; 26.3 ; 28.4 ; 29.1 ; 31.1 ; 31.7 ; 36.0 ; 52.0$; 55.8; 57.8; 61.3; 64.4; 73.9; 80.6; 111.6; 121.1; 127.0; 128.1; 128.2; 128.6; 128.8; 129.5; 129.6; 131.0; 131.1; 137.7; 139.0; 139.2; 149.8; 155.5; 156.8. HR-ESI-MS: $633.3344\left(\mathrm{MH}^{+}\right.$, $\mathrm{C}_{37} \mathrm{H}_{49} \mathrm{~N}_{2} \mathrm{O}_{5} \mathrm{~S}^{+}$; calc. 633.3357).

